Green silver nanoparticles ameliorate oxidative stress and apoptosis induced by gamma irradiation in rat pancreas

 \bf{M} arzouqah Almudayni¹, Raghad Aljohani¹, Sahar Khateeb^{1*}, Mody Albalawi¹, Kouther Alharthany¹, Noha Hamed²

¹Department of Biochemistry, Faculty of Science, University of Tabuk, Tabuk 71491, Saudi Arabia

²Nuclear Research Centre, Egyptian Atomic Energy Authority (EAEA), Cairo, Egypt

**Corresponding author's email:* skhateeb@ut.edu.sa

Received: 14 October 2024 / Accepted: 26 November 2024 / Published Online: 13 January 2025

Abstract

Radiation-related toxicity is a major concern for certain tissues and organs in radiation oncology practice. In abdominal tumor radiation treatment, the pancreas is particularly sensitive to radiation and should be considered at risk. The primary causes of acute pancreatitis after exposure to gamma radiation are oxidative damage and reactive oxygen species (ROS). The purpose of this study is to assess the efficacy of matcha silver nanoparticles (M-AgNPs) in mitigating oxidative stress and apoptosis induced by gamma radiation in the pancreas of female rats. Rats were exposed to 6 Gy of gamma radiation and subsequently administered an oral treatment with matcha (M) or M-AgNPs (10 ml/kg/day) for 14 days. We examined apoptotic markers such as caspase 3, B-cell lymphoma-2 (BCL2), and B-cell lymphoma-2-associated protein X (BAX) to evaluate their impact on cell survival. Additionally, the study investigated the modulation of antioxidants, glutathione S-transferases (GST), and malondialdehyde (MDA). The findings indicated that the administration of M-AgNPs for two weeks postradiation exposure is more efficacious in diminishing lipid peroxidation and suppressing apoptotic indicators compared to conventional M treatments. M-AgNPs significantly ($p < 0.05$) reduced the elevation of MDA and demonstrated a considerable ($p < 0.05$) increase in GST. Moreover, it exhibited a markedly elevated level ($p <$ 0.05) of BCL-2 and a significantly decreased level of Bax and caspase-3 ($p < 0.05$) in comparison to irradiated rats. The results of the histopathological investigations showed a notable enhancement in the histological characteristics of pancreatic tissue. In conclusion, the finding indicated that the AgNPs synthesized from matcha could potentially mitigate the adverse effects of radiation exposure. Further investigation is required to elucidate specific molecular pathways and their long-term consequences.

Keywords: Gamma radiation, Matcha, Silver nanoparticles, Oxidative stress, Apoptosis, Pancreas

How to cite this article:

Almudayni M, Aljohani R, Khateeb S, Albalawi M, Alharthany K and Hamed N. Green silver nanoparticles ameliorate oxidative stress and apoptosis induced by gamma irradiation in rat pancreas. Asian J. Agric. Biol. 2025: 2024213. DOI: <https://doi.org/10.35495/ajab.2024.213>

This is an Open Access article distributed under the terms of the Creative Commons Attribution 4.0 License. [\(https://creativecommons.org/licenses/by/4.0\)](https://creativecommons.org/licenses/by/4.0), which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

Introduction

While radiation therapy (RT) is a highly efficacious method for addressing malignant tumors, it has the potential to inflict damage on adjacent healthy tissues and give rise to severe adverse effects (Rezaeyan et al., 2016). The pancreas is indeed very sensitive to irradiation and should be considered an organ at risk during radiation treatment for abdominal tumors. Previous studies by Gemici et al. (2013) established that abdominal irradiation induces substantial structural and functional alterations in pancreatic tissue. Also, Wydmanski et al. (2016) discovered in a prior investigation that abdominal irradiation resulted in exocrine functional loss of pancreatic tissue in a subset of patients.

In RT, reactive oxygen species (ROS) are produced through the radiolysis of water in extracellular environments, and these highly reactive entities are detrimental to both tumor cells and nearby normal tissues (Zou et al., 2017). Moreover, radiation can provoke endogenous ROS generation in mitochondria and modify mitochondrial membrane permeability, thereby enhancing ROS production (Kim et al., 2015). Elevated levels of ROS can disrupt intracellular redox homeostasis (Kim et al., 2019), leading to cellular dysfunction and lipid peroxidation, which indicate oxidative stress and elevated malondialdehyde (MDA) levels (Wang et al., 2017; Zhang and Gurunathan, 2016). Moreover, when the level of ROS-induced DNA damage exceeds the cell's ability to repair, the cell activates the apoptotic pathway. Consequently, it is imperative to implement strategies to mitigate oxidative damage and its consequences resulting from radiation exposure.

Antioxidants are crucial for reducing oxidative damage resulting from radiation exposure. Cells possess an effective antioxidant system, comprising enzymes and non-enzymatic substances, that mitigates the detrimental effects of free radicals. Antioxidants can effectively suppress and/or treat chronic illnesses by obstructing or decelerating the interactions of biomolecules with free radicals through electron transfer, hence impeding the oxidative process. Antioxidant defense includes various mechanisms. These include delaying or inhibiting the production of free radicals; scavenging free radicals; converting free radicals into less toxic compounds; delaying the formation of secondary toxic active species; interrupting chain propagation reactions; enhancing the endogenous antioxidant defense system through synergistic interactions with other antioxidants; and chelating metal ions (Adwas et al., 2019; Costa et al., 2021).

Several disorders linked to oxidative stress are predominantly handled and treated according to the antioxidant properties of plant extracts. Furthermore, the notable antioxidant characteristics demonstrated by some nanomaterials present a compelling possibility to develop innovative regimens with enhanced and tailored efficacy. For instance, studies have demonstrated that the capacity of gold, silver, and selenium nanoparticles to eliminate redox-active radicals can mitigate oxidative stress (Saad et al., 2017; Sood and Chopra, 2017; Thilagavathi et al., 2016).

According to a recent study by Ali et al. (2024), coating or doping nanoparticles with alternative materials reduces their negative effects, enhances stability, and reduces agglomeration. The green biosynthesis of metal nanoparticles utilizing medicinal plant extracts is a significant research focus due to its relevance in various fields, particularly in medicine administration (Chaudhuri et al., 2016). It is a suitable alternative to chemical methods due to its cost efficiency and environmental friendliness (Njue et al., 2020). Plants include many bioactive compounds that help the synthesis of metal nanoparticles by serving as reducing and stabilizing agents (Begum et al., 2022).

Silver nanoparticles (AgNPs) have been the subject of extensive research due to their cost-effectiveness and various advantageous properties, including optical, antimicrobial, anticancer, and antioxidant (Vijayan et al., 2018; Suresh et al., 2014). As a consequence, the fabrication of particles on a nanoscale has emerged as a promising therapeutic alternative (Bejarbaneh et al., 2023). Polyphenolic compounds are natural compounds found in some plants, such as matcha green tea (M). M is the unfermented and finely ground green tea powder (Koláčková et al., 2020). It has many antioxidant and anti-inflammatory compounds (Kochman et al., 2020). The electron-donating capability of the phenolic compounds in matcha, which contributes to its higher total phenolic content, enables the reduction of silver ions to nanoscale silver particles. Recent studies indicate that nanoparticles

created from bioactive phytochemicals have superior beneficial and effective qualities compared to conventional herbal medications (Habeeb et al., 2022). Consequently, the purpose of this study is to assess the efficacy of the biosynthesized M-AgNPs in mitigating oxidative stress and apoptosis induced by gamma radiation in the pancreatic tissue of female rats.

Material and Methods

Material

Matcha green tea (Camellia sinensis) was bought from ILEAF NATURLS in the United States. Silver nitrate (99.0%) and polyvinylpyrrolidone-stabilized AgNPs were procured from Sigma Aldrich. All supplementary chemicals were of analytical grade and procured from reputable commercial providers.

Preparation and characterization of M-AgNPs

According to our previous research methodology (Hamed et al., 2023), M-AgNPs were created. M was used in the biogenic synthesis and green sonochemical approach to make AgNPs. M‐AgNPs were characterized using a variety of methods, including dynamic light scattering (DLS), high-resolution transmission electron microscopy (HR‐TEM), Fourier-transform infrared spectroscopy (FTIR), and thermogravimetric analysis (TGA).

Experimental animal groups

Rats were exposed to a single dose of whole-body gamma radiation (γ -Rad) at a rate of 0.33 Gy/min. γ -Rad at the designated dose of 6 Gy has the potential to stimulate apoptosis by inducing oxidative stress (Hamed and Hammad, 2023). In this study, we used 36 Wistar albino female rats, each weighing between 180 and 200 g, and divided them into six groups: the control group (C) received deionized water orally. The M group was given 10 ml/kg/day of M orally for a period of 14 consecutive days (Ninsiima et al., 2023). The MN group received 10 ml/kg/day of M-AgNPs orally over a 14-day period. Rats in the R group received a single dose of 6 Gy of whole-body ℽ-Rad, after which they received no treatment for the duration of the experiment. After 24 hours of receiving a single dose of whole-body γ -Rad (6 Gy), the MR group administered 10 ml/kg/day of M orally for 14 consecutive days. The MNR group received 10 ml/kg/day of M-AgNPs orally for 14 consecutive days after receiving a single dose of whole-body γ -Rad (6) Gy).

Sample preparation

Twenty-four hours after the final administration of M and M-AgNPs, the rats were euthanized for sample collection. Pancreas tissues were swiftly collected from the rats in various groups (10% wt/v) and homogenized in a Teflon apparatus using a cold PBS solution containing 0.16 mg/ml heparin. The homogenates underwent centrifugation at 4,000 rpm for 15 minutes at 4 °C. The transparent supernatant was employed to assess biochemical parameters subsequent to centrifugation.

Oxidative stress evaluation

Oxidative stress was measured using commercial kits (Bio-Diagnostic Company). Assessment of lipid peroxidation by measuring MDA. MDA was assessed via its reaction with thiobarbituric acid (TBA), resulting in the formation of colorful thiobarbituric acid reactive products (TBARs), which were quantified spectrophotometrically at 532 nm. MDA concentrations were measured in nmol/g of tissue. The GST kit assay quantifies overall GST activity (including cytosolic and microsomal) by assessing the conjugation of 1-chloro-2,4-dinitrobenzene (CDNB) with reduced glutathione. The GST activity was determined at 340 nm in the sample and expressed in U/gm of tissue.

Apoptotic markers evaluation

ELISA kits were used to determine caspase-3 (Cat. No. CSB-E09785r; CUSABIO, Wuhan, China), BAX (Cat. No. SEB631Ra; Cloud-Clone Corp., USA), and BCL-2 (Cat. No. E-EL-R0648; Elabscience®, USA)

in pancreas homogenate samples, according to the manufacturer's instructions.

Histopathological examination

After fixation with 10% formol saline, pancreatic tissue was washed and dehydrated in alcohol. Dehydrated specimens were cleaned in xylene, sealed in paraffin blocks, and sectioned at 4-6 µm thickness. Before histological analysis under an electric light microscope (Bancroft et al., 2013), tissue sections were deparaffinized with xylol and stained with H&E. Pancreatic tissue H&E sections were evaluated for the severity of apoptosis at ten distinct fields and

1.6 1.8 2

MDA (nmol/gm tissue)

200x magnification using a three-point scale: (parenchyma apoptosis—0: absent, 1: focal (<5%), 2: sub-lobular \langle <20%), 3: lobular (>20%). The analysis was performed in a blinded manner (Heindl et al., 2015).

Statistical analysis

Results were presented as mean values accompanied by standard error. The data were statistically examined utilizing one-way ANOVA, followed by Tukey's HSD multiple comparisons as a post-hoc test to identify significant differences across groups. The statistically significant level was set at $P < 0.05$. The software SPSS statistical version 20 (SPSS® Inc., USA) was used for all statistical analyses.

Results

Impact of M and M-AgNPs on oxidative stress

The results of our study indicated that the concentrations of MDA and GST did not significantly $(p > 0.05)$ rise in the M group (77.76 \pm 1.41 and 1.51) \pm 0.14, respectively) when compared to the control group (74.96 \pm 2.19 and 1.24 \pm 0.05 for MDA and GST). Furthermore, we detected no significant decrease ($p > 0.05$) in the MDA concentrations

between the MN group (62.60 ± 4.05) and control groups. However, a substantial elevation in GST ($p <$ 0.05) was noted in the MN group (1.77 ± 0.06) in comparison to the control groups. On the contrary to the control group, the radiation group demonstrated a significant reduction in GST levels $(0.29 \pm 0.02; p \leq$ 0.05) and a considerable increase in MDA levels $(123.66 \pm 3.57; p \le 0.05)$ (Figures 1A and 1B).

The data presented in Figure 1A clearly indicates that the MNR group significantly $(98.48 \pm 2.18; p \le 0.05)$ mitigated the rise in MDA when compared to the irradiated group. However, the MR (111.93 ± 2.58) and R groups showed no significant decrease ($p >$ 0.05). In addition, MDA concentrations were significantly ($p \leq 0.05$) lower in the MNR group compared to the MR group. The GST concentration in the MR (0.82 \pm 0.04; p < 0.05) and MNR (0.98 \pm 0.02; $p < 0.05$) groups was considerably greater than the irradiated group, as displayed in Figure 1B. This study's results indicated that M-AgNPs enhance cellular antioxidant defenses and exhibit significant antioxidant efficiency in mitigating gamma radiationinduced oxidative stress in the pancreas.

ab

 $\mathbf{\hat{c}}$

a

C M M N R M R MNR

(A)

a

ab

 $\frac{b}{b}$

b

b

Impact of M and M-AgNPs on apoptotic markers

The present investigation assesses the impact of M and M-AgNPs on pancreatic apoptotic markers. We evaluated the levels of BCL-2, Bax, and caspase-3 in all experimental groups. The M group exhibited significant increases in Bax (194.56 \pm 2.92; p < 0.05) and caspase 3 levels $(0.49 \pm 0.047; p < 0.05)$, while BCL2 levels (0.87 ± 0.03) did not increase significantly when compared to the control group $(110.56 \pm 0.72; 0.34 \pm 0.003; 0.83 \pm 0.04$ for Bax, caspase 3, and BCL2). Additionally, the MN group significantly elevated Bax levels $(240.90 \pm 7.87; p \le$ 0.05) compared to the control group, while caspase 3

did not significantly increase (0.47 ± 0.006) , and BCL2 (0.80 ± 0.02) levels did not significantly decrease.

Furthermore, in contrast to the control group, the radiation group exhibited a notable decrease in BCL2 levels $(0.49 \pm 0.04; p < 0.05)$, while Bax and caspase-3 levels significantly increased (514.73 \pm 6.65, p < 0.05; 0.81 ± 0.051 , $p < 0.05$, respectively) (Figure 2). Additionally, the findings of our study indicated that the MR and MNR groups exhibited a substantially higher concentration of BCL-2 (1.16 \pm 0.11, p < 0.05; 1.36 \pm 0.04, p < 0.05, respectively) than the radiation group. In addition, the MR and MNR groups demonstrated a significantly reduced concentration of Bax level $(383.56\pm4.31, p < 0.05; 417.2\pm7.60, p <$ 0.05) and caspase-3 level $(0.52 \pm 0.023, p \le 0.05)$; 0.45 ± 0.03 , $p < 0.05$) compared to the radiation group. Moreover, it is notable that in the treated groups, the M treatment demonstrated a significant decrease in Bax levels compared to M-AgNPs.

Figure 2. Impact of M and M-AgNPs on apoptotic markers: (A) BCL2, (B) Bax, and (C) Caspase 3 in the pancreas. Values were reported as means \pm SE (n = 6). a = significant vs. the control group at p < 0.05; b = significant vs. the irradiation group at $p < 0.05$; c = significant between the MR and MNR groups at $p < 0.05$.

Histopathological finding

A section of pancreatic tissue from the control, M, and MN animal groups revealed a pancreas with a normal architecture. Both the exocrine and endocrine tissues

of the pancreas exhibited typical histological characteristics. The lobules of the pancreas varied in size and morphology. The cellular components of the islets of Langerhans were organized normally, and the acinar structure contained typical proteinous eosinophilic materials. The pyramidal cells

comprising the acinar cells had acidophilic cytoplasm at their apex. The exocrine components of the pancreas are densely packed with acinar cells and organized into small lobules; they receive a score of 0 (figure 3a-b-c). The radiation-exposed animal group exhibited apoptosis of a subset of Langerhans cells. The acinar epithelial lining of the pancreas exhibited degenerative alterations accompanied by the absence of typical lobular architecture. Certain acinar cells exhibited vacuolation accompanied by nucleus pyknosis of score 3 (figure 3d). The animal group MR exhibited a significant quantity of Langerhans's cells undergoing apoptosis, with eosinophilic apoptotic bodies

Asian Journal of Agriculture and Biology

interspersed among them. Loss of normal lobular architecture accompanied by variable degenerative changes in the pancreatic acini; certain acinar cells exhibited vacuolation accompanied by pyknosis of their nuclei (figure 3e). In contrast, the histological appearance of the pancreatic lobules and islets of Langerhans improved notably in the animal group MNR. A subset of acinar cells exhibited pyknotic nuclei that were intensely stained, while the vast majority of acinar cells displayed vesicular nuclei. A few numbers of apoptotic Langerhans's cells were observed at score 1 (figure 3f).

Figure 3. Photomicrograph of pancreatic tissue section showing: (a,b,c) normal architecture of Langerhans's islets and acini; arrow (d) numerous numbers of apoptotic Langerhans's cells; arrow (e) greater numbers of apoptotic Langerhans's cells arrow (f): Few numbers of apoptotic Langerhans's cells arrow (H&EX 200).

Discussion

In radiation oncology, radiation-related toxicity is a crucial clinical consideration. The pancreas, due to its close proximity to the stomach and duodenum, may be unintentionally exposed to radiation during irradiation of the gastro-duodenal region (Zucca et al., 2020) or through total body irradiation. So, this study aims to investigate the potential impact of biosynthesized M-AgNPs in mitigating oxidative stress and apoptosis

induced by γ -radiation in the pancreatic tissue of female rats.

Contact with ionizing radiation (IR) produces ROS like hydroxyl radicals, superoxide, singlet oxygen, and hydrogen peroxide, which react with cellular components (Deng et al., 2019), causing morphological, metabolic, and cytotoxic changes due to oxidative stress and ROS, which damage lipids, proteins, and DNA (Hashim et al., 2020; Kamran et al.,

2021). Furthermore, Alattar et al. (2022) indicated that oxidative stress induces inflammation. In the present study, whole-body irradiation with a single radiation dose of 6 Gy elevated the MDA levels of pancreatic tissues. This aligns with research by Rezaeyan et al. (2016), who demonstrated that IR induces ROS within cells via water radiolysis, which subsequently assaults the fatty acids in the cell membrane and causes lipid peroxidation. Our results are also in line with those of other studies that used animal models and found that irradiation led to a significant rise in pancreatic MDA levels (Olgaç et al., 2006).

Contrarily, after irradiation, MDA levels were considerably lower in the M-AgNPs treatment (MNR group) than in the irradiated group. Nevertheless, the MR and R groups exhibited no substantial difference. Furthermore, the MNR group had substantially lower MDA concentrations than the MR group. This demonstrates that the M-AgNPs substantially reduced the increase in MDA, which correlates with oxidative stress and cellular damage. The results of this study are consistent with those of Guntur et al. (2018). Furthermore, a prior study indicated that AgNPs synthesized from green tea exhibited enhanced scavenging activity relative to green tea alone (Ruchikaa and Sehgala, 2020); this increase in scavenging efficacy may be ascribed to the increased surface area. The coating of polyphenolic residues on the surface of AgNPs may augment the interaction and ability of polyphenols in M to donate hydrogen to free radicals.

IR can adversely affect the antioxidant system; glutathione S-transferase (GST) is a crucial component of the cellular antioxidant system and plays key roles in preserving cellular homeostasis (Singh and Reindl, 2021). In the current investigation, GST levels markedly diminished in pancreatic tissues 14 days following irradiation. The present findings align with previous research indicating that IR might adversely affect the antioxidant system, leading to diminished levels and activity of antioxidant enzymes (Zhu et al., 2019).

However, following irradiation, the M or M-AgNPs treatment resulted in an increase in GST levels, indicating enhanced antioxidant defense mechanisms. Our results align with prior research indicating that plant-derived nanoparticles, such as silver, augmented the antioxidant activity of the respective molecules in the extract (Duman et al., 2016; Kanipandian et al., 2014). Moreover, a study by Al-Shmgani et al. (2017) indicates that AgNPs demonstrate antioxidant potential. Furthermore, our results indicated that the antioxidant activity of M-AgNPs surpassed that of matcha alone. Our findings correspond with those of Abdel-Aziz et al. (2014), who noted that plant extracts containing AgNP exhibited elevated amounts of total phenolic compounds and total flavonoids in comparison to the plant extract utilized independently. Nonetheless, certain studies indicate no alteration or a reduction in ROS following AgNP treatment (Qian et al., 2015; Gallorini et al., 2016; Pereira et al., 2018). The study's findings indicated that M-AgNPs can augment cellular antioxidant defenses and show considerable antioxidant effectiveness in mitigating gamma radiation-induced oxidative stress in the pancreas.

Apoptosis, a eukaryotic mechanism of cell death regulated by genetics, facilitates the controlled elimination of cells in order to preserve homeostasis and normal development (Singh et al., 2019). The cell survival and death are regulated by the equilibrium of pro-apoptotic and the anti-apoptotic BCL2 family proteins. The proteins in the BCL-2 family are divided into two groups: those that promote survival (BCL-2) and those that promote cell death (BAX) (Luna‐Vargas and Chipuk, 2016). Radiation-induced oxidative stress can initiate cell death by activating the mitochondriadependent apoptotic machinery. Apoptotic cell death involves the progressive activation of many cysteinedependent aspartate-directed proteases, commonly referred to as caspases. Furthermore, Chung et al. (2015) indicated that γ -radiation induces cell death via apoptosis and ROS generation. Elevated ROS levels trigger apoptosis by regulating the phosphorylation and ubiquitination of BCL2 family proteins, leading to the overexpression of pro-apoptotic genes (e.g., BAX) and the downregulation of anti-apoptotic genes (e.g., BCL2) (Li et al., 2004).

Our findings indicated that radiation caused a considerable reduction in BCL2 levels, whereas Bax and caspase-3 levels markedly rose. This finding is consistent with Changizi et al. (2021), who demonstrated that radiation markedly downregulated the BCL2 gene while upregulating the BAX and CASP3 genes. The significant decrease in Bcl-2 expression corresponds with prior research demonstrating the downregulation of this antiapoptotic protein following radiation exposure

(Rajabathar et al., 2023). In addition, the substantial increase in Bax expression highlights the proapoptotic signaling, which is consistent with studies linking elevated Bax levels to radiation-induced apoptosis (Mukherjee et al., 2022). Moreover, it was noted that heightened caspase-3 expression resulted from augmented oxidative stress caused by radiation (Li et al., 2015).

Conversely, our study's findings revealed that the MR and MNR groups demonstrated a significantly elevated level of BCL-2 compared to the radiation group. Furthermore, the MR and MNR groups had a markedly decreased level of Bax and caspase-3. M-AgNPs may inhibit apoptosis by diminishing lipid membrane peroxidation and oxidative stress, which are critical mediators of apoptosis. The distinctive phytochemical features of M-AgNPs, characterized by a high concentration of antioxidants such as catechins, along with their physicochemical attributes that augment biological activity, may facilitate their capacity to suppress apoptosis. Moreover, the combination of matcha's antioxidant-rich phytochemicals with the powerful properties of AgNPs may produce a synergistic effect, whereby the whole therapeutic outcome exceeds the individual effects of each component.

Conclusion

Radiation-related toxicity is a major concern for certain tissues and organs in radiation oncology practice. So, this study aims to investigate the potential impact of M-AgNPs in mitigating oxidative stress and apoptosis induced by γ-radiation in the pancreatic tissue of female rats. The results showed that administering M-AgNPs for two weeks after being exposed to six gray gamma radiation is more effective in reducing lipid peroxidation and suppressing apoptotic indicators than the conventional M treatment. The findings indicate that M-AgNPs could potentially act as effective agents in mitigating damage resulting from ionizing radiation exposure. M-AgNPs' unique phytochemical characteristics and their physicochemical characteristics enhance biological activity. Moreover, the combination of matcha's antioxidant-rich phytochemicals with the powerful properties of AgNPs may produce a synergistic effect, whereby the whole therapeutic outcome exceeds the individual effects of each component. Further investigation is required to elucidate specific molecular pathways and their longterm consequences.

Acknowledgment

The authors greatly thank and acknowledge the University of Tabuk, Kingdom of Saudi Arabia (KSA), for its support.

Disclaimer: None

Conflict of Interest: None

Source of Funding: This research has been supported by the University of Tabuk, Project number (S-1444-0064), University of Tabuk, Tabuk, Saudi Arabia.

Ethical Approval Statement: The ethics research committee of the National Center for Radiation and Technology of the Egyptian Atomic Energy Authority revised the protocol and granted its sanction (REC-NCRRT-10A/22).

Contribution of Authors

Khateeb S: Conceptualized study and supervised the research work, acquired funds, interpreted data and edited the final draft of manuscript.

Hamed NS: Designed the experiment and analyzed the data.

Aljohani R, Almudayni M, Albalawi M & Alharthany K: Prepared the initial draft of the manuscript, analyzed and interpreted data.

References

Abdel-Aziz MS, Shaheen MS, El-Nekeety AA and Abdel-Wahhab MA, 2014. Antioxidant and antibacterial activity of silver nanoparticles biosynthesized using Chenopodium murale leaf extract. J. Saudi Chem Soc. 18(4): 356- 363.

<https://doi.org/10.1016/j.jscs.2013.09.011>

Adwas AA, Elsayed A and Azab AE, 2019. Quwaydir, F.A. Oxidative stress and antioxidant mechanisms in human body. J. Appl. Biotechnol. Bioeng. 6: 43–47. [http://dx.doi.org/10.15406/jabb.2019.06.0017](http://dx.doi.org/10.15406/jabb.2019.06.00173) [3](http://dx.doi.org/10.15406/jabb.2019.06.00173)

Alattar A, Alvi AM, Rashid S, Hussain N, Gul M, Ikram M, Khalil AAK, Alshaman R, Shah FA, Li S and Li J, 2022. RETRACTED: Carveol ameliorates mercury-induced oxidative stress, neuroinflammation, and neurodegeneration in a mouse brain. <https://doi.org/10.1016/j.neuro.2022.08.006>

- Ali A, Saeed S, Hussain R, Saif MS, Waqas M, Asghar I, Xue X and Hasan M, 2024. Exploring the impact of silica and silica-based nanoparticles on serological parameters, histopathology, organ toxicity, and genotoxicity in Rattus norvegicus. Appl Surf Sci Adv. 19: 100551. <https://doi.org/10.1016/j.apsadv.2023.100551>
- Al-Shmgani HS, Mohammed WH, Sulaiman GM and Saadoon AH, 2017. Biosynthesis of silver nanoparticles from Catharanthus roseus leaf extract and assessing their antioxidant, antimicrobial, and wound-healing activities. Artif Cells Nanomed Biotechnol. 45(6): 1234-1240. [https://doi.org/10.1080/21691401.2016.1220](https://doi.org/10.1080/21691401.2016.1220950) [950](https://doi.org/10.1080/21691401.2016.1220950)
- Bancroft JD, Stevens A and Turner DR, 2013. Theory and practice of histological techniques. 4th Ed. Churchill Livingstone, Edinburgh, London, Melbourne, New York.
- Begum SJP, Pratibha S, Rawat JM, Venugopal D, Sahu P, Gowda A, Qureshi KA and Jaremko M, 2022. Recent advances in green synthesis, characterization, and applications of bioactive metallic nanoparticles. Pharmaceuticals. 15: 455. doi:10.3390/ph15040455
- Bejarbaneh M, Rahimi S, Nasiri M, Maivan AM, Ghasemian R, Davoudi A, Dashtmiani W, Sarai MDJP, Ghasabeh ZN, Kouchesfahani SS, Aghajani S, Ghasemipour T and Salehzadeh A, 2023. Cytotoxic effect of NiFe2O4@ Ag nanoparticle on adenocarcinoma gastric cell line (AGS) and assessment of the expression of CASP8, BAX, NRF2, and BCL2 genes. Gene Rep. 33: 101811.

<https://doi.org/10.1016/j.genrep.2023.101811>

Changizi V, Azadbakht O, Ghanavati R, Behrouj H, Motevaseli E and Khanzadeh P, 2021. Effect of Lactobacillus species on apoptosis-related genes BCL2, BAX, and caspase 3 in the testes *Asian Journal of Agriculture and Biology*

of gamma-irradiated rats. Rev Assoc Med Bras. 67: 1581-1585. <https://doi.org/10.1590/1806-9282.20210634>

- Chaudhuri SK, Chandela S and Malodia L, 2016. Plant mediated green synthesis of silver nanoparticles using Tecomella undulata leaf extract and their characterization. Nano Biomed Eng. 8(1): 1-8. <http://dx.doi.org/10.5101/nbe.v8i1.p1-8>
- Chung DM, Kim JH and Kim JK, 2015. Evaluation of MTT and Trypan Blue assays for radiationinduced cell viability test in HepG2 cells. Int. J. Radiat. Res. 13(4): 331. [https://api.semanticscholar.org/CorpusID:561](https://api.semanticscholar.org/CorpusID:56154742) [54742](https://api.semanticscholar.org/CorpusID:56154742)
- Costa M, Losada-Barreiro S, Paiva-Martins F and Bravo-Díaz C, 2021. Polyphenolic Antioxidants in Lipid Emulsions: Partitioning. Effects and Interfacial Phenomena. Foods. 10: 539. <https://doi.org/10.3390/foods10030539>
- Deng S, Shanmugam MK, Kumar AP, Yap CT, Sethi G and Bishayee A, 2019. Targeting autophagy using natural compounds for cancer prevention and therapy. Cancer. 125(8): 1228-1246. <https://doi.org/10.1002/cncr.31978>
- Duman F, Ocsoy I and Kup FO, 2016. Chamomile flower extract-directed CuO nanoparticle formation for its antioxidant and DNA cleavage properties. Mater Sci Eng C Mater Biol **Appl. 60:** 333-338. <https://doi.org/10.1016/j.msec.2015.11.052>
- Gallorini M, Di Giacomo V, Di Valerio V, Rapino M, Bosco D, Travan A, Di Giulio M, Di Pietro R, Paoletti S, Cataldi A and Sancilio S, 2016. Cell-protection mechanism through autophagy in HGFs/S. mitis co-culture treated with Chitlac-nAg. J Mater Sci Mater Med. 27: 1-11. [https://doi.org/10.1007/s10856-016-](https://doi.org/10.1007/s10856-016-5803-587) [5803-587](https://doi.org/10.1007/s10856-016-5803-587)
- Gemici C, Sargin M, Uygur-Bayramicli O, Mayadagli A, Yaprak G, Dabak R and Kocak M, 2013. Risk of endocrine pancreatic insufficiency in patients receiving adjuvant chemoradiation for resected gastric cancer. Radiother Oncol.

107(2): 195-199. <https://doi.org/10.1016/j.radonc.2013.04.013>

- Guntur SR, Kumar NS, Hegde MM and Dirisala VR, 2018. In vitro studies of the antimicrobial and free-radical scavenging potentials of silver nanoparticles biosynthesized from the extract of Desmostachya bipinnata. Anal Chem Insights. 13: 1177390118782877. <https://doi.org/10.1177/1177390118782877>
- Habeeb SA, Hammadi AH, Abed D and Al-Jibouri LF, 2022. Green synthesis of metronidazole or clindamycin-loaded hexagonal zinc oxide nanoparticles from Ziziphus extracts and its antibacterial activity. Pharmacia. 69(3): 855– 864. [http://dx.doi.org/10.3897/pharmacia.69.e910](http://dx.doi.org/10.3897/pharmacia.69.e91057)

[57](http://dx.doi.org/10.3897/pharmacia.69.e91057)

- Hamed NS and Hammad HB, 2023. The Protective Role of Red Beetroot (Beta Vulgaris L.) Peel Extract against Gamma Irradiation Induced Hepatic Apoptosis in Rats. Arab J. Nucl. Sci. Appl. 56(5): 69-79. [https://ajnsa.journals.ekb.eg/article_317892.h](https://ajnsa.journals.ekb.eg/article_317892.html) [tml](https://ajnsa.journals.ekb.eg/article_317892.html)
- Hamed NS, Taha EF and Khateeb S, 2023. Matcha‐ silver nanoparticles reduce gamma radiation‐ induced oxidative and inflammatory responses by activating SIRT1 and NLRP‐3 signaling pathways in the Wistar rat spleen. Cell Biochem Funct. 41(8): 1115- 1132. <https://doi.org/10.1002/cbf.3844>
- Hashim AM, Alharbi BM, Abdulmajeed AM, Elkelish A, Hozzein WN and Hassan HM, 2020. Oxidative stress responses of some endemic plants to high altitudes by intensifying antioxidants and secondary metabolites content. Plants. $9(7)$: 869. <https://doi.org/10.3390/plants9070869>
- Heindl M, Tuennemann J, Sommerer I, Mössner J and Hoffmeister A, 2015. Loss of Bace1 in mice does not alter the severity of caerulein induced pancreatitis. PLoS One. 10(5): e0125556. <https://doi.org/10.1371/journal.pone.0125556>
- Kamran M, Wang D, Alhaithloul HAS, Alghanem SM, Aftab T, Xie K, Lu Y, Shi C, Sun J, Gu W, Xu P and Soliman MH, 2021. Jasmonic acid-mediated enhanced regulation of

oxidative, glyoxalase defense system and reduced chromium uptake contributes to alleviation of chromium (VI) toxicity in choysum (Brassica parachinensis L.). Ecotoxicol Environ Saf. 208: 111758. <https://doi.org/10.1016/j.ecoenv.2020.111758>

- Kanipandian N, Kannan S, Ramesh R, Subramanian P and Thirumurugan R, 2014. Characterization, antioxidant and cytotoxicity evaluation of green synthesized silver nanoparticles using Cleistanthus collinus extract as surface modifier. Mater. Res. Bull. 49: 494-502. [https://doi.org/10.1016/j.materresbull.2013.0](https://doi.org/10.1016/j.materresbull.2013.09.016) [9.016](https://doi.org/10.1016/j.materresbull.2013.09.016)
- Kim W, Lee S, Seo D, Kim D, Kim K, Kim E, Kang J, Seong KM, Youn H and Youn B, 2019. Cellular stress responses in radiotherapy. Cells. 8(9): 1105. <https://www.mdpi.com/2073-4409/8/9/1105>
- Kim W, Youn H, Kang C and Youn B, 2015. Inflammation-induced radioresistance is mediated by ROS-dependent inactivation of protein phosphatase 1 in non-small cell lung cancer cells. Apoptosis. 20:1242–1252. <https://doi.org/10.1007/s10495-015-1141-1>
- Kochman J, Jakubczyk K, Antoniewicz J, Mruk H and Janda K, 2020. Health benefits and chemical composition of matcha green tea: A review. Molecules. 26(1): 85. <https://doi.org/10.3390/molecules26010085>
- Koláčková T, Sumczynski D, Zálešáková L, Šenkárová L, Orsavová J and Lanczová N, 2020. Free and bound amino acids, minerals and trace elements in matcha (Camellia sinensis L.): A nutritional evaluation. J. Food Compos. Anal. 92: 103581. [https://api.semanticscholar.org/CorpusID:225](https://api.semanticscholar.org/CorpusID:225252973) [252973](https://api.semanticscholar.org/CorpusID:225252973)
- Li B, Gao Y, Rankin GO, Rojanasakul Y, Cutler SJ, Tu Y and Chen YC, 2015. Chaetoglobosin K induces apoptosis and G2 cell cycle arrest through p53-dependent pathway in cisplatinresistant ovarian cancer cells. Cancer lett. 356(2): 418-433. <https://doi.org/10.1016/j.canlet.2014.09.023>
- Li D, Ueta E, Kimura T, Yamamoto T and Osaki T, 2004. Reactive oxygen species (ROS) control

Asian Journal of Agriculture and Biology

the expression of Bcl-2 family proteins by regulating their phosphorylation and ubiquitination. Cancer Sci. 95(8): 644-50. [https://doi.org/10.1111/j.1349-](https://doi.org/10.1111/j.1349-7006.2004.tb03323) [7006.2004.tb03323.](https://doi.org/10.1111/j.1349-7006.2004.tb03323)

- Luna‐Vargas MP and Chipuk JE, 2016. The deadly landscape of pro‐apoptotic BCL‐2 proteins in the outer mitochondrial membrane. FEBS J, 283(14), 2676-2689. <https://doi.org/10.1111/febs.13624>
- Mukherjee S, Dutta A and Chakraborty A, 2022. The cross-talk between Bax, Bcl2, caspases, and DNA damage in bystander HepG2 cells is regulated by γ-radiation dose and time of conditioned media transfer. Apoptosis. 27(3- 4):184-205. [https://doi.org/10.1007/s10495-](https://doi.org/10.1007/s10495-022-01713-4) [022-01713-4](https://doi.org/10.1007/s10495-022-01713-4)
- Ninsiima HI, Eze ED, Ssekatawa K, Nalugo H, Asekenye C, Onanyang D, Munanura EI, Ariong M, Matama K, Zirintunda G, Mbiydzenyuy NE, Ssempijja F, Afodun AM, Mujinya R, Usman IM, Asiimwe OH, Tibyangye J and Kasozi KI, 2023. Green tea silver nanoparticles improve physiological motor and cognitive function in BALB/c mice during inflammation. Heliyon. 9(3): e13922. [https://doi.org/10.1016/j.heliyon.2023.e1392](https://doi.org/10.1016/j.heliyon.2023.e13922) [2](https://doi.org/10.1016/j.heliyon.2023.e13922)
- Njue WM, Kithokoi JK, Mburu J, Mwangi H and Swaleh S, 2020. Green sonochemical synthesis of silver nanoparticles using Adansonia digitata leaves extract and evaluation of their antibacterial potential. Eur J Adv Chem Res. 1(2): 1‐7.
- Olgaç V, Erbil Y, Barbaros U, Öztezcan S, Kaya H, Bilge H, Güler S and Toker G, 2006. The efficacy of octreotide in pancreatic and intestinal changes: radiation-induced enteritis in animals. Dig Dis Sci. 51: 227-232. <https://doi.org/10.1007/s10620-006-3113-3>
- Pereira LC, Pazin M, Franco-Bernardes MF, da Cunha Martins JrA, Barcelos GRM, Pereira MC, Mesquita JP, Rodrigues JL, Barbosa F Jr and Dorta DJ, 2018. A perspective of mitochondrial dysfunction in rats treated with silver and titanium nanoparticles (AgNPs and TiNPs). J Trace Elem Med Biol. 47: 63-69. <https://doi.org/10.1016/j.jtemb.2018.01.007>
- Qian Y, Zhang J, Hu Q, Xu M, Chen Y, Hu G, Zhao M and Liu S, 2015. Silver nanoparticleinduced hemoglobin decrease involves alteration of histone 3 methylation status. Biomaterials. 70: 12-22. [https://doi.org/10.1016/j.biomaterials.2015.0](https://doi.org/10.1016/j.biomaterials.2015.08.015) [8.015](https://doi.org/10.1016/j.biomaterials.2015.08.015)
- Rajabathar JR, Al-Lohedan H, Arokiyaraj S, Mohammed F, Al-Dhayan DM, Faqihi NA and Al-Saigh H, 2023. Herbal Melanin Inhibits Real-Time Cell Proliferation, Downregulates Anti-Apoptotic Proteins and Upregulates Pro-Apoptotic p53 Expression in MDA-MB-231 and HCT-116 Cancer Cell Lines. Medicina (Kaunas). 59(12): 2061. <https://doi.org/10.3390/medicina59122061>
- Rezaeyan A, Haddadi GH, Hosseinzadeh M, Moradi M and Najafi M, 2016. Radioprotective effects of hesperidin on oxidative damages and histopathological changes induced by Xirradiation in rats heart tissue. J Med Phys. 41(3): 182-191. <https://doi.org/10.4103/0971-6203.189482>
- Ruchikaa MV and Sehgala A, 2020. Green tea derived silver nanoparticles boosted the antioxidant potential of greengreen tea. Plant Arch. 20(2): 3839‐3843.
- Saad AM, Abdel‐Aleem AAH, Ghareeb MA, Hamed MM, Abdel‐Aziz MS and Hadad AH, 2017. In vitro antioxidant, antimicrobial and cytotoxic activities and green biosynthesis of silver & gold nanoparticles using Callistemon citrinus leaf extract. J Appl Pharm Sci. 7(6): 141‐149. <https://dx.doi.org/10.7324/JAPS.2017.70620>
- Singh R, Letai A and Sarosiek K, 2019. Regulation of apoptosis in health and disease: the balancing act of BCL-2 family proteins. Nat Rev Mol Cell Biol. 20(3): 175-193. <https://doi.org/10.1038/s41580-018-0089-8>
- Singh RR and Reindl KM, 2021. Glutathione Stransferases in cancer. Antioxidants. 10(5): 701. <https://doi.org/10.3390/antiox10050701>
- Sood R and Chopra DS, 2017. Improved yield of green synthesized crystalline silver nanoparticles with potential antioxidant activity. Int Res J Pharm. 8(4): 100-104. <https://doi.org/10.7897/2230-8407.080457>
- Suresh G, Gunasekar PH, Kokila D, Prabhu D, Dinesh
- D, Ravichandran N, Ramesh B, Koodalingam A and Vijaiyan Siva G, 2014. Green synthesis of silver nanoparticles using Delphinium denudatum root extract exhibits antibacterial and mosquito larvicidal activities. Spectrochim Acta A Mol Biomol Spectrosc. 127: 61-66. <https://doi.org/10.1016/j.saa.2014.02.030>
- Thilagavathi T, Kathiravan G and Srinivasan K, 2016. Antioxidant activity and synthesis of silver nanoparticles using the leaf extract of Limonia acidissima. Int J Pharm Bio Sci. 7(4): 201‐ 205.

[http://dx.doi.org/10.22376/ijpbs.2016.7.4.b20](http://dx.doi.org/10.22376/ijpbs.2016.7.4.b201-205) [1-205](http://dx.doi.org/10.22376/ijpbs.2016.7.4.b201-205)

- Vijayan R, Joseph S and Mathew B, 2018. Indigofera tinctoria leaf extract mediated green synthesis of silver and gold nanoparticles and assessment of their anticancer, antimicrobial, antioxidant and catalytic properties. Artif Cells Nanomed Biotechnol. 46(4): 861-871. [https://doi.org/10.1080/21691401.2017.1345](https://doi.org/10.1080/21691401.2017.1345930) [930](https://doi.org/10.1080/21691401.2017.1345930)
- Wang TY, Libardo MDJ, Angeles-Boza AM and Pellois JP, 2017. Membrane oxidation in cell delivery and cell killing applications. ACS Chem Biol. 12(5): 1170-1182. <https://doi.org/10.1021/acschembio.7b00237>
- Wydmanski J, Polanowski P, Tukiendorf A and Maslyk B, 2016. Radiation-induced injury of the exocrine pancreas after chemoradiotherapy for gastric cancer. Radiother Oncol. 118(3): 535-539. <https://doi.org/10.1016/j.radonc.2015.11.033>
- Zhang XF and Gurunathan S, 2016. Combination of salinomycin and silver nanoparticles enhances apoptosis and autophagy in human ovarian cancer cells: an effective anticancer therapy. Int J Nanomedicine. 11: 3655-3675. <https://doi.org/10.2147/IJN.S111279>
- Zhu N, Liu R, He LX, Mao RX, Liu XR, Zhang T, Hao YT, Fan R, Xu MH and Li Y, 2019. Radioprotective effect of walnut oligopeptides against gamma radiationinduced splenocyte apoptosis and intestinal injury in mice. Molecules. 24(8): 1582. <https://doi.org/10.3390/molecules24081582>
- Zou Z, Chang H, Li H and Wang S, 2017. Induction of reactive oxygen species: An emerging approach for cancer therapy. Apoptosis. 22:1321–1335. <https://doi.org/10.1007/s10495-017-1424-9>
- Zucca E, Arcaini L, Buske C, Johnson PW, Ponzoni M, Raderer M, Ricardi U, Salar A, Stamatopoulos K, Thieblemont C, Wotherspoon A and Ladetto M, 2020. Marginal zone lymphomas: ESMO Clinical Practice Guidelines for diagnosis, treatment and follow-up. Ann Oncol. 31(1): 17-29. <https://doi.org/10.1016/j.annonc.2019.10.010>